

## CONFIDENTIAL FINAL REPORT

**SPONSOR:** VIROBLOCK SA  
**SPONSOR'S REPRESENTATIVE:** Thierry Pelet, Ph.D.  
**STUDY TITLE:** ASSESSMENT OF VIRUCIDAL EFFECTIVENESS OF TREATED FABRICS VIA DIRECT CONTACT- Influenza A Virus (H1N1) Misting Study  
**STUDY IDENTIFICATION:** MicroBioTest Project No. 798-126 (refer to signed protocol)

| TEST AGENT NAME                  | LOT NO.                | DATE RECEIVED | DS NO. |
|----------------------------------|------------------------|---------------|--------|
| FFP2 Respirator                  | 31001                  | 10/26/15      | F872   |
| FFP2 Respirator                  | 31005                  | 10/26/15      | F873   |
| FFP2 Respirator                  | 31009                  | 10/26/15      | F874   |
| FFP2 Respirator                  | 31016                  | 10/26/15      | F875   |
| FFP2 Respirator (Control Fabric) | VB-DEV-12-FEB-2013-NAT | 10/26/15      | F876   |

**ACTIVE INGREDIENT(S):** NPJ03  
**CHALLENGE ORGANISM:** Influenza A Virus, strain: A/PR/8/34 (H1N1); Charles River Laboratories  
**HOST CELL LINE:** MDCK cells; ATCC CCL-34  
**DILUTION MEDIUM:** 1X Minimum Essential Medium (MEM) + 3.0 µg/mL Trypsin  
**VIRUS SUSPENSION MEDIUM:** 0.1X MEM  
**ORGANIC LOAD:** Not required  
**NEUTRALIZER/EXTRACTION:** 1X MEM + 1% Fetal Bovine Serum (FBS) + 0.5% HEPES + 1% NaHCO<sub>3</sub> + 1mM EDTA

**EXPOSURE (CONTACT) TIME(S):** 10 minutes

**EXPOSURE TEMPERATURE:** Ambient temperature (20±1°C)

**APPLICATION:** As applicable, virus (or virus suspension medium) was misted through a 2 in x 2 in sterile barrier onto pre-cut test and control masks using Nalgene Aerosol Spray Bottle (Fisher Cat. # 15-232-8; Nalgene Cat. # 2430-0200) from a distance of 3 – 6” for two pumps, one second per pumps.

**NUMBER OF REPLICATES:** Four wells per dilution

**INCUBATION TEMPERATURE:** 36±2°C with 5±1% CO<sub>2</sub>

**INCUBATION TIME:** 4 – 6 days (5 days actual)

**CALCULATION OF TITER:**

The 50% tissue culture infectious dose per mL (TCID<sub>50</sub>/mL) was determined using the Spearman-Kärber method using the following formula:

$$m = x_k + \left(\frac{d}{2}\right) - d \sum p_i$$

where:

- m = the logarithm of the titer relative to the test volume
- x<sub>k</sub> = the logarithm of the smallest dosage which induces infection in all cultures
- d = the logarithm of the dilution factor
- p<sub>i</sub> = the proportion of positive results at dilution i
- ∑p<sub>i</sub> = the sum of p<sub>i</sub> (starting with the highest dilution producing 100% infection)

The values were converted to TCID<sub>50</sub>/mL using a sample inoculum of 1.0 mL.

## RESULTS

Results are presented in Tables 1 – 6.

The Viral Load was determined in the following manner:

Viral Load ( $\text{Log}_{10} \text{TCID}_{50}$ ) = Titer ( $\text{Log}_{10} \text{TCID}_{50}/\text{mL}$ ) +  $\text{Log}_{10}$  [Volume (mL)]

The  $\text{Log}_{10}$  Reduction Factor (LRF) was calculated in the following manner:

$\text{Log}_{10}$  Reduction Factor = Initial Viral Load ( $\text{Log}_{10} \text{TCID}_{50}$ ) – Output Viral Load  
( $\text{Log}_{10} \text{TCID}_{50}$ )

The Mean Viral  $\text{Log}_{10}$  Reduction from n replicates was determined as follows:

Mean Viral  $\text{Log}_{10}$  Reduction =  $(\text{LRF}_1 + \text{LRF}_2 + \dots + \text{LRF}_n) / n$

Note: The LRF's was anti-logged prior to performing calculations

**RESULTS (Continued):**

**Table 1  
Titer Results**

| Sample  | Replicate | Titer (Log <sub>10</sub> TCID <sub>50</sub> /mL) | Volume (mL) | Viral Load (Log <sub>10</sub> TCID <sub>50</sub> ) |
|---|-----------|--|-------------|--|
| Virus Stock Titer control   |           | 6.50   | -           | -  |
| Volume application evaluation                                       |           | average volume of challenge: 0.4 mL per run      |             |  |
| Cell viability/media sterility control                              |           | no virus detected, cells viable; media sterile   |             |  |
| Liquid (no fabric) Control  | Rep. 1    | 6.25   | 40          | 7.85   |
|   | Rep. 2    | 6.25   | 40          | 7.85   |
|   | Average   |  |             | 7.85   |
| FFP2 Respirator (Control Fabric)<br>(Lot No.VB-DEV-12-FEB-2013-NAT) | Rep. 1    | 5.25   | 40          | 6.85   |
|   | Rep. 2    | 5.00   | 40          | 6.60   |
|   | Average   |  |             | 6.74   |
| FFP2 Respirator (Lot No. 31001) <sup>b</sup>                        | Rep. 1    | 2.25   | 40          | 3.85   |
|   | Rep. 2    | 1.75   | 40          | 3.35   |
|   | Average   |  |             | 3.67   |
| FFP2 Respirator (Lot No. 31005) <sup>b</sup>                        | Rep. 1    | 2.75   | 40          | 4.35   |
|   | Rep. 2    | 2.50   | 40          | 4.10   |
|   | Average   |  |             | 4.24   |
| FFP2 Respirator (Lot No. 31009) <sup>b</sup>                        | Rep. 1    | 3.00   | 40          | 4.60   |
|   | Rep. 2    | 2.50   | 40          | 4.10   |
|   | Average   |  |             | 4.42   |
| FFP2 Respirator (Lot No. 31016) <sup>b</sup>                        | Rep. 1    | 2.75   | 40          | 4.35   |
|   | Rep. 2    | 2.50   | 40          | 4.10   |
|   | Average   |  |             | 4.24   |

<sup>b</sup> Cytotoxicity observed at an undilute dilution.

**RESULTS (Continued):**

**Table 2**

**Neutralizer Effectiveness/Viral Interference and Cytotoxicity Controls – Lot No. 31001**

| Dilution of the Test Agent/Neutralizer Mixture | Neutralizer Effectiveness Control                       | Cytotoxicity Control     |
|--|---|--------------------------|
| Undilute                                       | cytotoxicity observed; viral CPE could not be evaluated | cytotoxicity observed    |
| 10 <sup>-1</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |
| 10 <sup>-2</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |

**Table 3**

**Neutralizer Effectiveness/Viral Interference and Cytotoxicity Controls – Lot No. 31005**

| Dilution of the Test Agent/Neutralizer Mixture | Neutralizer Effectiveness Control                       | Cytotoxicity Control     |
|--|---|--------------------------|
| Undilute                                       | cytotoxicity observed; viral CPE could not be evaluated | cytotoxicity observed    |
| 10 <sup>-1</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |
| 10 <sup>-2</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |

**Table 4**

**Neutralizer Effectiveness/Viral Interference and Cytotoxicity Controls – Lot No. 31009**

| Dilution of the Test Agent/Neutralizer Mixture | Neutralizer Effectiveness Control                       | Cytotoxicity Control     |
|--|---|--------------------------|
| Undilute                                       | cytotoxicity observed; viral CPE could not be evaluated | cytotoxicity observed    |
| 10 <sup>-1</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |
| 10 <sup>-2</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |

**Table 5**

**Neutralizer Effectiveness/Viral Interference and Cytotoxicity Controls – Lot No. 31016**

| Dilution of the Test Agent/Neutralizer Mixture | Neutralizer Effectiveness Control                       | Cytotoxicity Control     |
|--|---|--------------------------|
| Undilute                                       | cytotoxicity observed; viral CPE could not be evaluated | cytotoxicity observed    |
| 10 <sup>-1</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |
| 10 <sup>-2</sup>                               | virus detected in all inoculated wells                  | no cytotoxicity observed |

**RESULTS (Continued):**


**Table 6  
Viral Reduction Based on Liquid (no mask) Control**

| Test Article   | Input Load*<br>(Log <sub>10</sub> TCID <sub>50</sub> ) | Output Load*<br>(Log <sub>10</sub> TCID <sub>50</sub> ) | Log <sub>10</sub> Reduction |
|--|--|---|-----------------------------|
| FFP2 Respirator (Lot No. 31001)                                      | 7.85   | 3.67  | 4.18                        |
| FFP2 Respirator (Lot No. 31005)                                      |  | 4.24  | 3.61                        |
| FFP2 Respirator (Lot No. 31009)                                      |  | 4.42  | 3.43                        |
| FFP2 Respirator (Lot No. 31016)                                      |  | 4.24  | 3.61                        |
| FFP2 Respirator (Control Fabric)<br>(Lot No. VB-DEV-12-FEB-2013-NAT) |  | 6.74  | 1.11                        |

**CONCLUSION:**

VIROBLOCK SA's FFP2 Respirator Lot No.'s 31001, 31005, 31009 and 31016 were evaluated for the ability to inactivate Influenza A virus (H1N1). MicroBioTest personnel performed the inactivation procedure using the challenge viruses to spike the test material. Samples were tested and titrated by 50% tissue culture infectious dose (TCID<sub>50</sub>) endpoint assay using MDCK cells.

The log<sub>10</sub> viral reductions for the test materials are presented in Tables 6. All of the controls met the criteria for a valid test. These conclusions are based on observed data.

Study director:   
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11/24/2015  
Date